# SDM Practical

### SDM Pratical overview:



### SDM – Design your experiment

- <u>Think before starting</u>!
  - What do you want to know about the species?
  - How do you verify that?
- <u>Set an hypothesis</u>:
  - E.g. Climate change will decrease the area available for species X
    - Thus: HO: There is no change in the suitable area of species X HA: There is a decrease of the suitable area of species X
- <u>Draft the steps</u> to test said hypothesis:

Goto: <u>https://www.gbif.org/</u>





On the options on the left, make sure to only select occurrences with geographic information (and perhaps only human observation)

Does it have synonyms? <u>https://resolver.globalnames.org/</u> 27 synonyms for this bat species



#### Global Names resolution tools and services

Resolve lists of scientific names against known sources. This service parses incoming names, executes exact or fuzzy matching as required, and displays a confidence score for each match along with its identifier.

Paste Scientific Names, one on each

Rhinolophys euryale

on each	Bequito	
	Results	JSON XML
	Rhinolophys euryale	Number of matches: 27

- CSV comma separated variables
  - Comma's have different meanings:
  - EU (in general): Comma is a decimal separator
  - USA: point is the decimal separator
  - Also tab delimited (decimal can change here)
- R can read all types
- Correcting from csv1 to csv2
  - Open in notepad
  - Substitute , for ;
  - Substitute . for ,
- There are other methods (check the manual)

🥅 Rhinolophus_	euryale_csv0 - Blo	oco de notas 🛛 —		🔲 Rhinolophus_	euryale_csv1 - Bloco de nota	as	🗐 Rhinolophus	_euryale_csv2 - Bl	oco de notas
Ficheiro Editar	Formatar Ver	Ajuda		Ficheiro Editar	Formatar Ver Ajuda		Ficheiro Editar	Formatar Ver	Ajuda
"species"	"longitu	ude" "latitu	de"	"species","lo	ongitude","latitude	e"	species;long	gitude;latit	ude
"Rhinolophus	euryale"	10.261719	51.193676	"Rhinolophus	euryale",10.261719	9,51.193676	Rhinolophus	euryale;10,	261719;51,193676
"Rhinolophus	euryale"	5.566667	50.633333	"Rhinolophus	euryale",5.566667,	,50.633333	Rhinolophus	euryale;5,5	66667;50,633333
"Rhinolophus	euryale"	5.566667	50.633333	"Rhinolophus	euryale",5.566667,	,50.633333	Rhinolophus	euryale;5,5	66667;50,633333
"Rhinolophus	euryale"	20.167 48.617		"Rhinolophus	euryale",20.167,48	3.617	Rhinolophus	euryale;20,	167;48,617
"Rhinolophus	euryale"	20.75 48.5166	66	"Rhinolophus	euryale",20.75,48.	.516666	Rhinolophus	euryale;20,	75;48,516666
"Rhinolophus	euryale"	20.887191	48.495193	"Rhinolophus	euryale",20.887191	1,48.495193	Rhinolophus	euryale;20,	887191;48,495193
"Rhinolophus	euryale"	20.506927	48.467712	"Rhinolophus	euryale",20.506927	7,48.467712	Rhinolophus	euryale;20,	506927;48,467712
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	20.542033	48.460844	"Rhinolophus	euryale",20.542033	3,48.460844	Rhinolophus	euryale;20,	542033;48,460844
"Rhinolophus	euryale"	-0.29907	48.20928	"Rhinolophus	euryale",-0.29907,	,48.20928	Rhinolophus	euryale;-0,	29907;48,20928
"Rhinolophus	euryale"	-0.29907	48.20928	"Rhinolophus	euryale",-0.29907,	,48.20928	Rhinolophus	euryale;-0,	29907;48,20928
"Rhinolophus	euryale"	-0.29907	48.20928	"Rhinolophus	euryale",-0.29907,	,48.20928	Rhinolophus	euryale;-0,	29907;48,20928
"Rhinolophus	euryale"	16.917221	48.133888	"Rhinolophus	euryale",16.917221	1,48.133888	Rhinolophus	euryale;16,	917221;48,133888
"Rhinolophus	euryale"	20.749859	48.056655	"Rhinolophus	euryale",20.749859	9,48.056655	Rhinolophus	euryale;20,	749859;48,056655
"Rhinolophus	euryale"	20.749859	48.056655	"Rhinolophus	euryale",20.749859	9,48.056655	Rhinolophus	euryale;20,	749859;48,056655

ArcGIS, R, excel, etc, each of them might recognize numbers different so keep all files

- Maxent expects the data in CSV
  - Species name; longitude; latitude
    - MAKE SURE YOU HAVE ONE NAME PER SPECIES
    - When you download from GBIF, it usually brings multiple names
  - Yes, you can have multiple species in one file
    - Yes you can have multiple subsets of the same species in one file: e.g. amanita1, amanita2
      - They will all be modelled independently
- IMPORTANT STEP:
  - Load your CSV into a GIS and explore your #



	A	В	С	
1	species	longitude	latitude	
2	Rhinoloph	10,26172	51,19368	
3	Rhinoloph	5,566667	50,63333	
4	Rhinoloph	5,566667	50,63333	
5	Rhinoloph	20,167	48,617	
6	Rhinoloph	20,75	48,51667	
7	Rhinoloph	20,88719	48,49519	
8	Rhinoloph	20,50693	48,46771	
9	Rhinoloph	20,54203	48,46084	
10	Rhinoloph	20,54203	48,46084	
11	Rhinoloph	20,54203	48,46084	
12	Rhinoloph	20,54203	48,46084	
13	Rhinoloph	20,54203	48,46084	
14	Rhinoloph	20,54203	48,46084	
·				





In a GIS you can easily clean the obvious errors

- Further explore if the data "makes sense"
- When satisfied, export back to csv (check GIS notes)



Hum...



### Different sampling strategies



### Floating mushrooms

- Goto: <u>https://www.worldclim.org/</u>
  - Worldclim version 2: 1970~2000
    - Tif format (more GIS friendly)
    - Only "Present"
  - Worldclim version 1.4: 1960~1990
    - Bil format
    - Also a section for Future and Current
  - You can use either worldclim version but future scenarions are only possible on the previous version
  - Next:
    - Variable description
    - Coordinate system and resolution





#### WorldClim

WorldClim is a set of global climate layers (gridded climate data) with a spatial resolution of about 1  $\rm km^2$ . These data can be used for mapping and spatial modeling.

The new Version 2.0 is now available (current climate only --- more coming soon)

The old version is **Version 1.4**. For this version you can get data for past, current and future climates.

Read more

You can also directly download in R:

Example @ https://www.gis-blog.com/r-raster-data-acquisition/

### Main product of Wordclim dataset

• Monthly means of each of these estimates

variable	10 minutes	5 minutes	2.5 minutes	30 seconds
minimum temperature (°C)	tmin 10m	tmin 5m	tmin 2.5m	tmin 30s
maximum temperature (°C)	tmax 10m	tmax 5m	tmax 2.5m	tmax 30s
average temperature (°C)	tavg 10m	tavg 5m	tavg 2.5m	tavg 30s
precipitation (mm)	prec 10m	prec 5m	prec 2.5m	prec 30s
solar radiation (kJ m <sup>-2</sup> day <sup>-1</sup> )	srad 10m	srad 5m	srad 2.5m	srad 30s
wind speed (m s <sup>-1</sup> )	wind 10m	wind 5m	wind 2.5m	wind 30s
water vapor pressure (kPa)	vapr 10m	vapr 5m	vapr 2.5m	vapr 30 s

#### Coordinate system: World Geodetic System 84(WGS84)



### These are secondary products that "sumarize" the main products

variable	10 minutes	5 minutes	2.5 minutes	30 seconds	
Bioclimatic variables	bio 10m	bio 5m	bio 2.5m	bio 30s	

BIO1 = Annual Mean Temperature BIO2 = Mean Diurnal Range (Mean of monthly (max temp - min temp)) BIO3 = Isothermality (BIO2/BIO7) (\* 100) BIO4 = Temperature Seasonality (standard deviation \*100) BIO5 = Max Temperature of Warmest Month BIO6 = Min Temperature of Coldest Month BIO7 = Temperature Annual Range (BIO5-BIO6) BIO8 = Mean Temperature of Wettest Quarter BIO9 = Mean Temperature of Driest Quarter BIO10 = Mean Temperature of Warmest Quarter BIO11 = Mean Temperature of Coldest Quarter BIO12 = Annual Precipitation BIO13 = Precipitation of Wettest Month BIO14 = Precipitation of Driest Month BIO15 = Precipitation Seasonality (Coefficient of Variation) BIO16 = Precipitation of Wettest Quarter BIO17 = Precipitation of Driest Quarter BIO18 = Precipitation of Warmest Quarter BIO19 = Precipitation of Coldest Quarter

#### Resolution at the equator!

Conversion to m explained in: <u>https://en.wikipedia.org/wiki/Decimal\_degrees</u> X (m) = 30(s) \* Perimeter@equator/360(°) = 927.66m @ equator AKA 1km layer Download the 5 minute layer! (10km @ equator)

2050					
GCM	code	гср26	гср45	гср60	гср85
ACCESS1-0 (#)	AC		tn, tx, pr, bi		tn, tx, pr, bi
BCC-CSM1-1	BC	tn, tx, pr, bi			
CCSM4	CC	tn, tx, pr, bi			
CESM1-CAM5-1-FV2	CE		tn, tx, pr, bi		
CNRM-CM5 (#)	CN	tn, tx, pr, bi	tn, tx, pr, bi		tn, tx, pr, bi
GFDL-CM3	GF	tn, tx, pr, bi	tn, tx, pr, bi		tn, tx, pr, bi
GFDL-ESM2G	GD	tn, tx, pr, bi	tn, tx, pr, bi	tn, tx, pr, bi	
GISS-E2-R	GS	tn, tx, pr, bi			
HadGEM2-AO	HD	tn, tx, pr, bi			
Haddewiz SC	HG		tn, tx, pr, bi		tn, tx, pr, bi
HadGEM2-ES	HE	tn, tx, pr, bi			
INMCM4	IN		tn, tx, pr, bi		tn, tx, pr, bi
IPSL-CM5A-LR	IP	tn, tx, pr, bi			
MIROC-ESM-CHEM (#)	MI	tn, tx, pr, bi			
MIROC-ESM (#)	MR	tn, tx, pr, bi			
MIROC5 (#)	MC	tn, tx, pr, bi			
MPI-ESM-LR	MP	tn, tx, pr, bi	tn, tx, pr, bi		tn, tx, pr, bi
MRI-CGCM3	MG	tn, tx, pr, bi			
NorESM1-M	NO	tn, tx, pr, bi			

0050

#### Year the scenario represents

Greenhouse gas scenarios: four representative concentration pathways (RCPs) Time periods: 2050 (average for 2041-2060) and 2070 (average for 2061-2080) Variables:

tn - monthly average minimum temperature (degrees C \* 10) tx - monthly average maximum temperature (degrees C \* 10) pr - monthly total precipitation (mm) bi - 'bioclimatic' variables



Download the data corresponding to the scenarios you want to test

#### Notice: you must use the same variables for present and forecasting

- Be organized!
  - Think of folders as a "database"
  - If you use R script: 00 SettingUpWorkEnvironment.R you will get an environment like mine
- Unzip each .zip file to the appropriate folders
  - My case: Future data to Future folder, Present data to Present folder
- First step: subsetting the data to your Area of Interest
  - <u>Alternative 1: Clip your AOI based on the range of your presence data</u>
  - Alternative 2: Clip your AOI based on a polygon shapefile created in Arcmap
  - Alternative 3: Create your own extent
- You should train your model in an AOI and then predict back to the entire planet
  - If you want to see the future in a specific AOI, you can also clip the future layers
  - We show that on the tutorial but it's optional



- Overall procedure:
  - 1. Load the entire group of rasters in the present
  - 2. Load an object (occurrence points or a shapefile) to crop the rasters to na AOI
  - 3. Select the variables of interest using an ecological criteria
  - 4. Test the selected data for Autocorrelation and VIF
    1. If >.7 or VIF >10, then you have to remove somethig
  - 5. Save all the data to a new folder in .asc format
    - 1. Maxent needs it!
  - 6. Start-up maxent, load the data, set the settings and run



#### The direction of the slash is significant!!!

Single \ - tells Rm next character is special Double: \\ - Tells R, next character is a backslash Forward: / - recognized as a backslash

For R you must use: \\ or / when using a folder path or you will have an error

```
Error: unexpected input in "\"
> setwd("C:\Practical\")
Error: '\P' is an unrecognized escape in character string starting ""C:\P"
> setwd("C:/Practical/")
> |
```

Originally: (Bio1, bio10 ... Bio 19, bio 2... bio9)

But we want: Bio1, bio2, .... Bio19

them

#### Now you repeat the same for the scenarios you want to use

```
#loading future variables is a bit more complicated due to the subfolders
27
    path.fut.26 <- list.files("./Future/he26bi50/",pattern=".tif",full.names = T)</pre>
28
    path.fut.45 <- list.files("./Future/he45bi50/",pattern=".tif",full.names = T)</pre>
29
    path.fut.60 <- list.files("./Future/he60bi50/",pattern=".tif",full.names = T)</pre>
30
    path.fut.85 <- list.files("./Future/he85bi50/",pattern=".tif",full.names = T)</pre>
31
32
33
    stk.fut.26 <- stack(path.fut.26)</pre>
    stk.fut.45 <- stack(path.fut.45)</pre>
34
35
    stk.fut.60 <- stack(path.fut.60)
    stk.fut.85 <- stack(path.fut.85)</pre>
36
37
    #checks the order of the layers loaded and renames them to bioXX, each XX repr
38
    #a bioclimatic laver
39
    names(stk.present)
40
    names(stk.present) <- c("Bio01","Bio02","Bio03","Bio04",</pre>
41
42
                              "Bio05", "Bio06", "Bio07", "Bio08",
43
                              "Bio09", "Bio10", "Bio11", "Bio12",
44
                              "Bio13", "Bio14", "Bio15", "Bio16",
                              "Bio17", "Bio18", "Bio19")
45
46
```

This is to rename the variables of the present period to the bioclim naming – <u>if you do it now, it helps you later when</u> <u>selecting the variables. Everything will be in proper order</u>

In the case of the future variables they are often loaded out of order,

#### you should check in your case!!

names(stk.fut.45) <- list.of.names

names(stk.fut.60) <- list.of.names
names(stk.fut.85) <- list.of.names</pre>

78

79

80

81

```
[1] "./Future/he26bi50/he26bi501.tif" "./Future/he26bi50/he26bi5010.tif" "./Future/he26bi50/he26bi5011.tif
                                                                                                            [4] "./Future/he26bi50/he26bi5012.tif" "./Future/he26bi50/he26bi5013.tif" "./Future/he26bi50/he26bi5014.tif
48 names(stk.fut.26)
                                                                                                                "./Future/he26bi50/he26bi5015.tif" "./Future/he26bi50/hg26bi5016.tif" "./Future/he26bi50/he26bi5017.tif
49
    names(stk.fut.45)
                                                                                                           [10] "./Future/he26bi50/he26bi5018.tif" "./Future/he26bi50//he26bi5019.tif" "./Future/he26bi50/he26bi502.tif"
[13] "./Future/he26bi50/he26bi503.tif" "./Future/he26bi504.tif" "./Future/he26bi505.tif"
   names(stk.fut.60)
50
                                                                                                                "./Future/he26bi50/he26bi506.tif" "./Future/he26bj50/he26bi507.tif" "./Future/he26bi50/he26bi508.tif"
51
   names(stk.fut.85)
                                                                                                           [19] "./Future/he26bi50/he26bi509.tif"
52
    path.fut.26
53
    #the easiest way to is just to re-load the variables again with the proper order
   path.fut.26 <- list.files("./Future/he26bi50/",pattern=".tif",full.names = T[c(1,12:19,2:11)]
54
55 path.fut.45 <- list.files("./Future/he45bi50/",pattern=".tif",full.names = T)[c(1,12:19,2:11)]</pre>
56 path.fut.60 <- list.files("./Future/he60bi50/",pattern=".tif",full.names = T)[c(1,12:19,2:11)]
                                                                                                                          Now the John Position
    path.fut.85 <- list.files("./Future/he85bi50/",pattern=".tif",full.names = T)[c(1,12:19,2:11)]
57
58
59
    stk.fut.26 <- stack(path.fut.26)</pre>
60
    stk.fut.45 <- stack(path.fut.45)</pre>
   stk.fut.60 <- stack(path.fut.60)</pre>
61
62
    stk.fut.85 <- stack(path.fut.85)</pre>
63
64
    #check if the∨ loaded fine
    names(stk.fut.26)
65
66
   names(stk.fut.45)
67
    names(stk.fut.60)
                                                                                  > path.fut.26
68
    names(stk.fut.85)
                                                                                                                           "./Future/he26bi50/he26bi502.tif" "./Future/he26bi50/he26bi503.tif"
                                                                                   [1] "./Future/he26bi50/he26bi501.tj/f"
69
                                                                                   [4] "./Future/he26bi50/he26bi504.tif"
                                                                                                                           "./Future/he26bi50/he26bi505.tif" "./Future/he26bi50/he26bi506.tif"
                                                                                   [7] "./Future/he26bi50/he26bi507.tif"
70
    #and then rename them easily
                                                                                                                           "./Future/he26bi50/he26bi508.tif" "./Future/he26bi50/he26bi509.tif"
                                                                                       "./Future/he26bi50/he26bi5010.tif" "./Future/he26bi50/he26bi5011.tif" "./Future/he26bi50/he26bi5012.tif"
71
     list.of.names <- c("Bio01","Bio02","Bio03","Bio04",
                                                                                  [10]
                                                                                  [13] "./Future/he26bi50/he26bi5013.tif" "./Future/he26bi50/he26bi5014.tif" "./Future/he26bi50/he26bi5015.tif"
72
                           "Bio05", "Bio06", "Bio07", "Bio08",
                                                                                                                           "./Future/he26bi50/he26bi5017.tif" "./Future/he26bi50/he26bi5018.tif"
                                                                                  [16]
                                                                                       "./Future/he26bi50/he26bi5016.tif"
                           "Bio09", "Bio10", "Bio11", "Bio12",
73
                                                                                  [19] "./Future/he26bi50/he26bi5019.tif
74
                           "Bio13", "Bio14", "Bio15", "Bio16",
75
                           "Bio17","Bio18","Bio19")
76
77
    names(stk.fut.26) <- list.of.names
```

> path.fut.26

When everything is in proper order, we can name it – <u>Maxent likes it</u> when variables for projections share names with the training variables

- Creating an extent object
  - We will clip to the extent of the area where we have occurrences
    - For a polygon area, the exercise is the same (ish).

#### • First load the occurrence data:

```
names(stk.fut.26)
82
83 # Load species occurrence file
84 # notice im using read.csv2, which expects a EU type of table. If you want to use the NA style,
85 #then you must switch read.csv2 with read.csv
86 #You can also use custom delimitrs
   sp <- read.csv2("./Occurrences/Rhinolophus_euryale_csv2.csv",header=T) #load csv of occurrence
87
88
   head(sp) #check table looks correct
89
   sp_shp <- sp #rename table</pre>
   coordinates(sp_shp) <- ~longitude+latitude #convert table to points shapefile
90
91
    proj4string(sp_shp) <- CRS("+proj=longlat +ellps=WGS84 +datum=WGS84 +no_defs")</pre>
92
93
                                                                           20
94 #Create bounding box around points
                                                                           45
    bbox <- extent(sp_shp) #create bounding box of points</pre>
95
   bbox <- bbox+2 #increase border so we do not truncate data
96
                                                                           40
97
    plot(stk.present$Bio01,ext=bbox+2)
                                                                           35
    plot(bbox, col='blue',add=T) #check if box surrounds points
98
    plot(sp_shp,add=T,pch=19,col='red') #add points
99
                                                                           30
```

60

Here i had a problem – some coordinate values

were wrong, if you have an error, eliminate those rows in Excel, save as csv, and repeat

20

40

### • Clipping the rasters:

- 101 #cropping the present data
- 102 stk.present.AOI.crop <- crop(stk.present,bbox) #clip to training area
- 103 #plotting the example
- 104 par(mfrow=c(1,2)) #sets the plotting area to a 1 line 2 columns set up
- 105 plot(stk.present\$Bio01,main="Original extent")
- 106 plot(stk.present.AOI.crop \$Bio01,main="Cropped extent")
- 107 par(mfrow=c(1,1)) #sets it back to 1 image per plot

### • The crop command clips all rasters

```
#now we can save them to another folder in a format
115
116
     #that maxent can read
117
     #saving the cropped present data in .asc format
118
     writeRaster(stk.present.AOI.crop,
                 "./Present_AOI/.asc",
119
120
                 overwrite=T.
121
                 bylayer=T,
122
                 suffix="names")
172
```



#### te PC > Windows (C:) > Practical > Present rdy Nome Data de modificação Tipo Tamanho Bio01.asc 22/11/2019 16:06 Ficheiro ASC 3 770 KB Bio02.asc 22/11/2019 16:06 Ficheiro ASC 3 769 KB Bio03.asc 22/11/2019 16:06 Ficheiro ASC 3 769 KB Bio04.asc Ficheiro ASC 22/11/2019 16:06 3 769 KB Bio05.asc 22/11/2019 16:06 Ficheiro ASC 3 750 KB

With what we have we could go to maxent, but first we need to check for autocorrelation effects in our data

- Now we repeat the same with the present data (world, cropped by future scenarios antartica is not there in the future data
  - And then you save it to the proper folder (check the script!)

```
108
109 #cropping the future data
110 stk.fut.26.AOI.crop <- crop(stk.fut.26,bbox)</pre>
111 stk.fut.45.AOI.crop <- crop(stk.fut.45,bbox)</pre>
112
     stk.fut.60.AOI.crop <- crop(stk.fut.60,bbox)</pre>
     stk.fut.85.AOI.crop <- crop(stk.fut.85,bbox)</pre>
113
114
115 #you can see the
116 par(mfrow=c(1,2))
     plot(stk.fut.26$Bio01)
117
     plot(stk.present$Bio01)
118
119
120 #cropping the present world data
     stk.present <- crop(stk.present,stk.fut.26)</pre>
121
1 - - -
```

- Remember autocorrelation and multicollinearity
- Pairwise correlation



- Model multicollinearity
  - Select n variables X1... Xn
  - Sucessively make a linear model:
    - X1 ~ X2...Xn
    - X2 ~ X1 + X3... Xn
    - IF any VIF > 10, remove said variable
  - Repeat above step with new model, excluding variable X that was removed.
  - Don't worry, R has a package for it

$$VIF_i = \frac{1}{1 - R_i^2}$$

### Pairwise correlation test

### • Pearson coefficient

21 ### the autocorrelation testing is important ONLY for the areas where
22 ### model is trained, so, for this section, we use only the cropped e
23

24 ### pairwise testing

2 V -

- 25 #first we convert the cropped raster to a data.frame
- 26 stk.present.AOI.crop <- na.omit(as.data.frame(stk.present.AOI.crop))</pre>
- 27 #now this stores the pearson correlation in a matrix
- 28 cor.tab <-cor(stk.present.AOI.crop)</pre>
- 29 #remember to change to write.csv if needed
- 30 write.csv2(cor.tab,"CorrelationTable\_AOI.csv")

A	В	С	D	E	F	G	н	- I -	J	K	L	M	N	0	Р	Q	R	S	Т	
	Bio01	Bio02	Bio03	Bio04	Bio05	Bio06	Bio07	Bio08	Bio09	Bio10	Bio11	Bio12	Bio13	Bio14	Bio15	Bio16	Bio17	Bio18	Bio19	
Bio01	1	0,686055	0,660273	-0,23315	0,875827	0,820743	0,071517	0,025992	0,844474	0,891814	0,913634	-0,56364	-0,44576	-0,63376	0,685389	-0,45453	-0,63032	-0,68202	-0,26329	
Bio02	0,686055	1	0,659006	0,111116	0,81381	0,293616	0,509326	-0,10757	0,650355	0,730932	0,506533	-0,60923	-0,51346	-0,63433	0,645142	-0,52374	-0,644	-0,64656	-0,37722	
Bio03	0,660273	0,659006	1	-0,64641	0,409402	0,724167	-0,28811	-0,22129	0,608793	0,356148	0,803072	-0,15062	-0,07727	-0,28211	0,561358	-0,07839	-0,26755	-0,35339	0,075049	
Bio04	-0,23315	0,111116	-0,64641	1	0,243434	-0,71377	0,909285	0,18432	-0,18764	0,228713	-0,60605	-0,40359	-0,41508	-0,25636	-0,14273	-0,42122	-0,28258	-0,19758	-0,44821	
Bio05	0,875827	0,81381	0,409402	0,243434	1	0,452432	0,539321	0,068535	0,764088	0,989195	0,614411	-0,75876	-0,64563	-0,76733	0,639536	-0,65728	-0,77509	-0,78853	-0,47539	
Bio06	0,820743	0,293616	0,724167	-0,71377	0,452432	1	-0,50698	-0,05363	0,682023	0,494753	0,971028	-0,14168	-0,05604	-0,27523	0,507805	-0,05796	-0,25494	-0,34915	0,090673	
Bio07	0,071517	0,509326	-0,28811	0,909285	0,539321	-0,50698	1	0,116884	0,094506	0,488915	-0,32306	-0,59959	-0,57111	-0,48176	0,13863	-0,58056	-0,50842	-0,43246	-0,5451	
Bio08	0,025992	-0,10757	-0,22129	0,18432	0,068535	-0,05363	0,116884	1	-0,28637	0,10579	-0,06057	-0,15776	-0,19362	0,020961	-0,22475	-0,19881	0,004904	0,186607	-0,36424	
Bio09	0,844474	0,650355	0,608793	-0,18764	0,764088	0,682023	0,094506	-0,28637	1	0,76456	0,774035	-0,48841	-0,36614	-0,64124	0,672729	-0,37387	-0,62398	-0,75172	-0,12991	
Bio10	0,891814	0,730932	0,356148	0,228713	0,989195	0,494753	0,488915	0,10579	0,76456	1	0,635333	-0,74352	-0,63235	-0,75114	0,608282	-0,64363	-0,75762	-0,77618	-0,46092	
Bio11	0,913634	0,506533	0,803072	-0,60605	0,614411	0,971028	-0,32306	-0,06057	0,774035	0,635333	1	-0,28708	-0,18652	-0,41157	0,611413	-0,19089	-0,39586	-0,47981	-0,0184	
Bio12	-0,56364	-0,60923	-0,15062	-0,40359	-0,75876	-0,14168	-0,59959	-0,15776	-0,48841	-0,74352	-0,28708	1	0,941355	0,843861	-0,42529	0,95245	0,871175	0,804919	0,844799	
Bio13	-0,44576	-0,51346	-0,07727	-0,41508	-0,64563	-0,05604	-0,57111	-0,19362	-0,36614	-0,63235	-0,18652	0,941355	1	0,651168	-0,17839	0,996067	0,683382	0,667598	0,896504	
Bio14	-0,63376	-0,63433	-0,28211	-0,25636	-0,76733	-0,27523	-0,48176	0,020961	-0,64124	-0,75114	-0,41157	0,843861	0,651168	1	-0,67518	0,668518	0,992771	0,909616	0,511984	
Bio15	0,685389	0,645142	0,561358	-0,14273	0,639536	0,507805	0,13863	-0,22475	0,672729	0,608282	0,611413	-0,42529	-0,17839	-0,67518	1	-0,19707	-0,67729	-0,61078	-0,07466	
Bio16	-0,45453	-0,52374	-0,07839	-0,42122	-0,65728	-0,05796	-0,58056	-0,19881	-0,37387	-0,64363	-0,19089	0,95245	0,996067	0,668518	-0,19707	1	0,699409	0,680928	0,903838	
Bio17	-0,63032	-0,644	-0,26755	-0,28258	-0,77509	-0,25494	-0,50842	0,004904	-0,62398	-0,75762	-0,39586	0,871175	0,683382	0,992771	-0,67729	0,699409	1	0,903737	0,550295	
Bio18	-0,68202	-0,64656	-0,35339	-0,19758	-0,78853	-0,34915	-0,43246	0,186607	-0,75172	-0,77618	-0,47981	0,804919	0,667598	0,909616	-0,61078	0,680928	0,903737	1	0,399641	
Bio19	-0,26329	-0,37722	0,075049	-0,44821	-0,47539	0,090673	-0,5451	-0,36424	-0,12991	-0,46092	-0,0184	0,844799	0,896504	0,511984	-0,07466	0,903838	0,550295	0,399641	1	

- Think of which variables are good for your species
- Confirm if there is more than 0.7 and smaller than -.07 pairwise correlation between them
- To many pairwise correlations will cause multicollinearity

	Var 1	Var 2	Va3
Var 1	Var 1 ~ var 1		
Var 2	Var2 ~var 1	Var 2 ~ var 2	
Var3	Var 3 ~ Var 1	Var 3 ~ var 2	

• For exemple, my first selection:

• Now I need to test for multicollinearity:

	Bio01	Bio04	Bio07	Bio12	Bio15	Bio19
Bio01	1	-0,23315	0,071517	-0,56364	0,685389	-0,26329
Bio04	-0,23315	1	0,909285	-0,40359	-0,14273	-0,44821
Bio07	0,071517	0,909285	1	-0,59959	0,13863	-0,5451
Bio12	-0,56364	-0,40359	-0,59959	1	-0,42529	0,844799
Bio15	0,685389	-0,14273	0,13863	-0,42529	1	-0,07466
Bio19	-0,26329	-0,44821	-0,5451	0,844799	-0,07466	1

#multicollinearity testing library(usdm) #e.g. i select Bio01; Bio04; Bio07; Bio 12; Bio 15 and bio 19 head(df.crop.stack) df.crop.stack.selection <- df.crop.stack[,c(1,4,7,12,15,19)] #select only the variables i am interested head(df.crop.stack.selection)

vif(df.crop.stack.selection, maxobservations=nrow(df.crop.stack.selection))

- Bio 04 and 07 are so strongly correlated that they affect the entire model I have to remove one
  - Bio 04: Temp. seasonality
  - Bio 07: Temp. anual range
  - I like seasonality more for Mediterranean climates

	Va	riables	VIF				
1		Bio01	3.623841				
2		Bio04	13.534628				
3		Bio07	13.308972				
4		Bio12	9.459776				
5		Bio15	2.617598				
6		Bio19	5.691570				

	Var	iables	VIF
1		B1001	3.285936
2		Bio04	1.934874
3		Bio12	9.448515
4		Bio15	2.331450
5		Bio19	5.659382

## Last step before modelling

### • Now we have:

- Selected the occurrence data we want to use
- Selected the variables we want to use and cropped them to our choosen study area
- What's missing?
  - Crop the future scenario data and export it to a folder

#Saving the cropped variables to the correct folder, in the .asc format

#we had already saved the cropped rast path2rst.HadGEM2ES\_RCP85 <- list.files("./Future/", full.names = T) path2rst.HadGEM2ES\_RCP85 #in this case the order of variables is changed, its better to keep everything in the #same order. It's easy to adapt the code to read in our preferred order #WARNING: this step might be different in your case, CHECK IT FIRST rst.fut.stack.crop <- crop(rst.fut.stack,bbox) names(rst.fut.stack.crop)

#now we can jus save the variables to a new folder #but first, we rename the variables to the same as the present vars names(rst.fut.stack.crop)<- names(rst.AOI.crop)</pre>

Think:

- If you train and project the model in one AOI you can't find new areas around the world where the species might be
- If you crop your future data, you will also not be able to predict anything regarding the world

# Maxent finally

Load the occurrences (beware of the type of CSV)

t tir	hallv		Load the enviromental variables – point						
	iany		to the folder						
	🙆 Maximum Entropy Species Distribution Modeling, Versio	on 3.4.1		×					
	Samples	Environr	nental layers						
	File rrences\Rhinolophus_euryale_csv1.c Browse	Directory/File C:\Practical\Presen	t_AOI Browse	9					
		Bio09	Continuous						
		BI010	Continuous		De-select all and then select				
		Bio12	Continuous		vour variables				
		Bio13	Continuous		your variables				
		Bio14	Continuous 🗸						
	Rhinolophus_euryale	✓ _Bio15	Continuous 🗸						
5V)			Continuous 💌	=					
		Bio17	Continuous 🗸	]					
		<b>Bio18</b>	Continuous 💌						
		✓ _Bio19	Continuous	-					
		Select all	Deselect all		Activate overything and				
	✓ Linear features		Create response curves						
	V Quadratic features		Make pictures of predictions		choose logistic output				
	Product features	Do jackknife	to measure variable importance [		0				
	Threshold features		Output format Logistic	-					
			Output file type asc	•					
	Output dire	ectory C:\Practical\Maxent\Results_v1	4 Browse	•					
	Projection	layers directory/file D:\Practical\Future	e_VVLD\he45bi50_VVLD Browse	•					
	Run	Settings	Help						
			Droce cottings	for out	ra antiona				
			riess settings	s ior ext	ra options				

Take note of the warnings you will get and let me know what you think they are x)

# Maxent finally

